Study of Collection Efficacy of Cell Separator Machine and Engraftment outcome of **Peripheral Blood HSCT**

Dr. Rima Kusumgar¹, Dr Sandip Shah², Dr. Manoj Shah³, Dr. Pragnesh Shah⁴

Abstract

To evaluate collection Efficacy of cell separator machines and post transplant outcome of stem cell therapy collected from peripheral blood in various haematological conditions. Present study of 170 patients who underwent autologus and allogenic stem cell transplant collected from peripheral blood of both adult and paediatric age groups from year 1999 to 2012. Total 308 procedures were done among which 72(23%) were on Cs 3000 plus and rest on Amicus cell separator. 218 procedures were autologus and 90 were allogenic procedures. Mononuclear cell collection protocols on cell separators were adopted for harvesting PBSCs with the help of aphaeresis catheter. Mean collection efficacy of the Amicus aphaeresis machine is 75% and median yield is 6.6×10^8 MNC/Kg body weight. A median of 5.6×10^6 CD 34+ cells per kg recipient body weight were collected. On CS 3000 plus average collection efficacy was 50 % and yield 5.54×10^8 MNC/Kg body weight. 2.1×10^6 CD 34+ cells per kg body weight were harvested in a median product volume 50 ml. Post transplant median time for neutrophilic and platelet recovery is 12 days and 16 days respectively. Only 2 patients of Aplastic anemia and 2 patients of thalassemia had primary graft rejection. 6 patients died due to GVHD grade IV. Other patients had low grade reversible GVHD. Collection of PBSCs using the Amicus cell separator allows adequate yields of MNCs and CD34+ cells. Though the values of MNC count and CD 34+ cells are lower in CS3000 plus, outcome of engraftment was not affected. Engraftment takes place in more than 95% of the patients. Stem cell transplant is boon to the patients where other treatment is not responding or available.

Key Words: Engraftment, graft versus host disease, mononuclear cell (MNC), Peripheral blood stem cell

¹Assistant Professor and In-charge Blood bank, ²Consultant Hematologist, ³ Professor & Head, dept. of Pathology, GCRI Medical college, Ahmedabad, ⁴Associate Professor and In-charge Blood bank, Government Medical College, Bhavnagar, Gujarat, India.

Corresponding author mail: rima_27@rediffmail.com

2013

Introduction: In the recent times, peripheral blood stem cells (PBSCs) have increasingly become a major source of hematopoietic stem cells for transplantation in patients with various hematological and oncological conditions and are being preferred over stem cells obtained from the bone marrow.¹

Traditionally, bone marrow was always the source of choice for stem cells in the treatment of various clinical conditions primarily because there are more progenitor cells, and presumably stem cells in the small volume of bone marrow than in the 5 litres of circulating blood in a human.² During the early attempts at using peripheral cells blood stem for transplantation, the challenge was to number address the very low of hematopoietic stem cells present in the circulating peripheral blood.^{2,3}

Later it was established that the number of circulating hematopoietic stem cells dramatically increased under various conditions, especially during the recovery from ablative phase following chemotherapy. This made the process of mobilization of these cells into circulation subsequent collection and their for transplantation more feasible.^{2,3}

Clinical results of peripheral blood stem cell transplantation (PBSCT) have shown benefits of earlier hematopoietic recovery, faster engraftment, lower morbidity, and greater cost-effectiveness compared with the conventional bone marrow transplantation.⁴In addition, the relative ease of obtaining large amounts of stem cells has made stem cell transplantation a option in the treatment of viable malignancies. The method also provides the flexibility of further increase of chemotherapy dose intensity in certain conditions.³

In recent years, there has been an increase in PBSCT performed for malignant conditions, both hematological and solid tumors. The number of PBSCTs has rapidly surpassed the number of bone marrow transplants (BMTs) performed in the autologous setting and PBSCT is also increasingly used in the allogenic setting.⁵

Optimum mobilization of stem cells into the peripheral circulation is the key to a successful PBSCT. The higher number of progenitor cells in mobilized compared with steady-state peripheral blood enables sufficient cell harvest with fewer collection sittings. Also, the benefits of hematopoietic enhanced recovery in PBSCT are only seen using mobilized collections. Clinically, mobilization

regimes consist of chemotherapy or hematopoietic growth factors (such as G-CSFs) or both.^{2,3}

After mobilization, PBSCs are collected by multiple apheresis procedures on separators.⁶ automated cell **PBSCs** represent a diverse group of cells with different functional capacities, but a majority of them express CD34 antigen that is usually used for their detection.⁶A significant increase in the number of PBSCTs and technological advancements has led to the development of new and more efficient apheresis devices, improving PBSC collections.⁶

Since the advent of automated cell separators, PBSC collection has been well established on the first generation of Cell Separators – CS-3000plus (Baxter-Fenwal) and Spectra (Cobe).^{1,37} The machines target collection of mononuclear cells (MNCs) from mobilized donors and patients. The MNCs include the CD34+ stem cells, which are the ones needed for the transplantation.³

As in the developed nations, PBSCT has been slowly gaining wider acceptance in India. Various Cancer Institute's and Hematology-Oncology set-ups in the country have recognized the benefits of using stem cells in various malignant and non-malignant conditions.

For CS-3000plus long, has been consistently and successfully used in many centres in India for PBSC collection and harvesting.^{5,7} Controlled product volume with high MNC and CD34+ yield as well superior collection efficiency has as always been achieved and documented collection on CSthrough PBSC 3000plus.^{6,7} Technology has led to the introduction of second generation cell separators designed to provide faster, comfortable and efficient collections(Ex. Amicus).⁶

Various international studies have demonstrated that the PBSCs collected by the Amicus device are viable after transfusion and capable of inducing a durable engraftment that was comparable with those published for other cell separators.^{5,6,7,8}

Additionally, the mean Amicus CD34+cell collection efficiency was better and platelet content in the stem cell products significantly lower than that of the CS-3000plus.^{6,7}

We have been using CS-3000plus since over 10 years for PBSC collection and have started using the Amicus since last 5 years. In the current paper, we compared the CD34+cell collections by Amicus and CS-3000plus both in autologous and allogeneic settings for adult and pediatric patients. The collection efficiencies and the engraftment outcomes of both the machines were studied.

Materials and Methods:

Patients:

The study included 170 patients of diverse hematological conditions of both adult and pediatric age groups from year 1999 to 2012. During first 6 years only 46 procedures (26 patients) were done. All were using CS 3000 plus only. Total 308 procedures were done among which 218(70.8%) autologus from were 109(64%) patients and 90(29.2%) were allogenic procedures from 71(41%) patients. 72(23%) of the procedures were done using CS 3000 plus (Baxter) and rest the procedures were done on of AMICUSTM (Baxter/Fenwal) Procedures were started only after taking informed consent and performing serological tests (HIV,HBsAg,HCV) of either patients or donors. Diagnosis wise defferentiation is shown in Chart I.

34(20%) patients of allogenic transplant were of paediatric age group.

The patients were mobilized with G-CSF (10ug/kg/day) following various chemotherapy regimens. Collections were begun at the discretion of the attending physician.

PBSC collection:

MNC protocols on both the cell separators were adopted for harvesting PBSCs with the help of apheresis catheter. In case of Pediatric age group, priming was done by anticoagulated cross-matched blood. Blood volume processed three times higher than the body volume. Per cycle volume kept 1000 ml if WBC count is more than 35000/ul and 1400 ml if WBC count is less than 35000/ul. Two pints of Normal calcium Saline with gluconate and magnesium sulphate running during process.

For collection on the CS-3000plus and Amicus cell separators, parameter settings as per the manufacturer's recommendations were used.

The PBSC product was analyzed for volume, WBCs, MNCs, and CD34+cell yields.

WBC, MNC and hematocrit were determined using an automated cell counter. MNC count was confirmed using a Leishman stain and manual count.

CD34+cell determination:

CD34 +cells were enumerated in the peripheral blood and apheresis component by flow cytometry.

In case of allogenic mismatched ABO group transplant RBCs were removed by adding HES (ratio 1:8).

Autologous stem cells were preserved by adding cryoprotectant DMSO and stored at -80 ⁰ C till transplantation. CD 34 count was done by Flow Cytometer.

Engraftment was reported by periodical check of blood counts and chimerism study. Days of engraftment were considered after infusion of CD34 + cells when absolute neutrophil count \geq 500/ul and platelets \geq 20000/ul for 3 consecutive days.

Results:

Results were studied for product out come in the form of volume, MNC/Kg body weight and CD 34+ cells/Kg body weight, mean collection efficiency of equipments, average processing time and engraftment results in the form of platelet and neutrophilic recovery and are shown in **Table 1.** No severe adverse effects were observed during or after the PBSC collection procedures other than mild hypocalcemic symptoms. Only 9(6%) of the total aphaeresis collections were interrupted due to venous access problems but it was solved after changing the inlet to out let and vice versa.

Only 2(1.17%) patients of Aplastic anemia and 2(1.17%) patients of thalassemia had primary graft rejection. 6 patients died due to GVHD grade IV. Other patients had low grade reversible GVHD.

Table 2 shows comparison of patient andcollection variables of two aphaeresisequipments used in our study.

Chart 1: Diagnosis of Patients

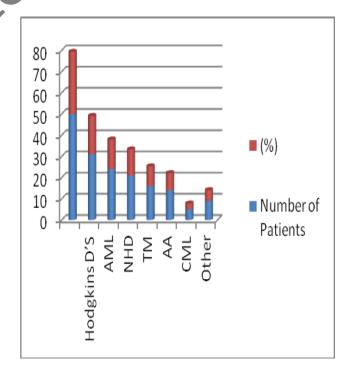


Table 1: Mean of Apheresis EquipmentProduct Data

Table 2: Comparison of Patient &Collection Variables

		Amicus	CS	1				AMICU
		Anneus	CS				CS 3000	
Volume	Adult	12,500	11,600(800	-				S
					Patients (number)		72	98
Processed		(9000-	0-13000)		Age	Adult	53.1(32-	53.4(32-
		15000)			(years)		<u>_68</u>)	68)
	Pedia	8300(43	81,00(4200-		(years)			,
	tric	00-	10000)			Pediatric	6.2(2.5-	6.5(3-
		9000)					14)	14)
Auticogulant		948(612	840(600-		Gender	Male	40(23.5%)	47(27.6)
Consumption		-1148)	1250)			Female	32(18.8%)	51(30%)
(ml.)		,	,		Body	Adult	66	65.2
Flow rate		50(38-62)	60(50-70)		Weight/			
		50(50 02)	00(30-70)		Kg	Pediatric	18.5	20.2
(ml/min)								
Separation time		5(4.2-5.6)	4.5(2.5-5)		Daninhana		20.0	40.0
(hrs)*					Periphera		38.2	40.0
Product Allo		205(88-	50(45-55)		$(*10^{3}/\text{ ul})$			
Volume		275)			Platelets	(*10 ³ /ul)	2.05	1.48
	Auto	220(95-			Periphera	al Hct (%)	28.4	33.2
		265)			- r			
MNC * 10 ⁸ /Kg* 6.6(1.17			5.6(1.5-6.5)					
		-8.7)						
CD 34 * 1	$0^{6}/Kg^{*}$	5.54	2.1					
NOTE: * p value < 0.05								

Table	3:	Comparison	with	Other
Studies	5			

Parameters			p-Value
MNC*10 ⁸	Our	6.6(n=124)	0.02
/Kg	study		
U U	Snyder	7.8(n=36)	
(Amicus)	et al.	/.o(ii 20)	
Noutrophilio		12(n-170)	0.01
Neutrophilic	Our	12(n=170)	0.01
Recovery	Study		
(Days)	Snyder	8.7(n=31)	
	et al.		
	our		
	study		0.015
	Yale	9.3(n=400)	
	et al.		
Platelete	Our	16(n=170)	0.009
Recovery	Study		
(Days)	Snyder	9.7(n=31)	
	et al.		
	our	10.9(n=40	0.002
	study	0)	
	Yale		
	et al.		
Collection	Our	75(n=124)	0.001
Efficacy of	Study		
Amicus	Yale	50(n=46)	
	et al.		

Discussion: A successful engraftment of the collected hematopoietic cells in a patient is based on optimum mobilization of stem cells into the peripheral blood and

an efficient collection of CD34+ cells during the harvest. A superior CD34+ collection efficiency improves CD34+ harvest yields and ensures sufficient cells to go ahead with the infusion and transplantation. The CD34+efficiency depend on various factors; significant among them is the use of the appropriate cell separator. In this study, we evaluated the performance of the Amicus Cell Separator of Fenwal for the first time in our institution. At the same time, we product compared yields and CD34+efficiency of two separators. The CS-3000plus, which had been using effectively since a long time and Amicus (both of Fenwal, Baxter) on two groups of donors – Allogeneic and Autologous, that did not differ significantly in terms of their age, gender, diagnosis, body weight, blood indices and initial CD34+ cell concentrations.

Our study has shown that AMICUS cell separator effectively collects MNCs. Successful collections were seen in adult as well as paediatric patients. The results are comparable and similar to various studies published earlier.^{3,4,5} The CD 34+ yields per harvest and of cumulative harvests in allogeneic donors were higher than autologous harvests.⁹ Collection of PBSCs using the Amicus cell separator allows adequate yields of MNCs and CD34+ cells. Though the values of MNC count and CD 34+ cells are lower in CS 3000 +, outcome of engraftment was not affected. Though the Amicus procedure takes little longer than the one on CS-3000plus, the platelet contamination in the product is lower than the CS-3000plus.^{5,9} The comparative results are in line with other comparative studies published in literature.^{4,6,7,10}

Engraftment times for allogenic and autologous PBSCs were also equivalent to data published in the literature.^{1,3,7} Because it is more efficient than the CS-3000plus, the Amicus separator would be more useful for collecting PBSCs from patients who are poor mobilizers and who have marginal circulating CD34+ cells.^{6,7}

As far as GVHD is concerned, the results are equivalent to stem cells collected from bone marrow. Engraftment takes place in more than 95% of the patients.

Table 3 shows comparison with various studies. In our study although the CD 34+ cells/Kg was slightly lower than other studies, our engraftment results were quite acceptable.

Conclusion: The CS 3000+ and Amicus separator are effective and efficient in the collection of a good yield of PBSCs

though Amicus collects CD34+ cells with greater efficiency and collects MNCs with higher yield than does the CS-3000 Plus, the engraftment outcome results are equivalent to stem cells collected from bone marrow. Stem cell transplant is a boon to those patients for whom other treatments are not effective or available.

References:

- 1. Ikeda K, Ohto H, Ogata K, et al. Automated programs for collection of mononuclear cellsand progenitor two cell separators cells by for peripheral blood progenitor cell transplantation: comparison by а randomized crossover study. Transfusion 2007; 47: 1234-1240 Baynes R, Hamm C, et al. Bone Marrow and Peripheral
 - BloodHematopoietic Stem Cell Transplantation:Focus on Autografting Clinical Chemistry 2000; 46:8(B) 1239–1251
- Ikeda K, Ohto H, Nemoto K, et al. Collection of MNCs and progenitor cells by two separators for PBPC transplantation: a randomized crossover trial. Transfusion 2003; 43: 814-819
- Moog R,Basu O, et al. Collection of Peripheral Progenitor Cells inPediatric Patients with a New Programmefor the Collection of Mononuclear Cells.

leukapheresis

Journal of Clinical Apheresis 2003; 18:111–1143

- Snyder E, Baril L, Min K, et al. In vitro collection and posttransfusion engraftment characteristics of MNCs obtained by using a new separatorfor autologous PBPC transplantation. Transfusion 2000; 40:961-967.
- Jeanne M, Bouzgarrou R, et al. Comparison of CD34+ cell collection on the CS-3000+ and Amicus blood cell separators. Transfusion 2003; 43: 1423-1427.
- Snyder E, O'Donnell L, Dengler T, et al. Ex vivo evaluation of PBMNCscollected with a new cell separator. Transfusion 2001;41:940-949
- Burgstaler E, Pineda A, et al. Hematopoietic Progenitor Cell Large VolumeLeukapheresis (LVL) on the Fenwal AmicusBlood Separator. Journal of Clinical Apheresis 2004; 19:103–111
- 9. Burgstaler E,Porrata L, et al. Use of Various Offset Settings in the Fenwal Amicusduring Hematopoietic
 Progenitor Cell Collection toIncrease
 Lymphocyte Yield and Reduce
 Cross-cellularContamination.
 - Journal of Clinical Apheresis 2010; 25:301–309
- 10. Schwella N, Movassaghi K, Scheding S, et al. Comparison oftwo

programs

for